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Complexation of vanadium(V) oxyanions with hexopyranose- and mannopyranoseuronic acid-containing polysaccharides: stereochemical considerations

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Abstract—Carbohydrates containing galactopyranosyl and mannopyranosyl units with vicinal *cis*-diols were treated with NaVO₃ in D₂O, and complexation was determined by ⁵¹V NMR spectroscopy. Me α-Galp, Me β-Galp (3,4-*cis*-diols), and Me α-Manp (2,3-*cis*-diol) complexed, but Me β-Manp barely did so. This low degree of complexation also occurred with a β-mannan containing alternate (1 \rightarrow 3)- and (1 \rightarrow 4)-linkages and an alginate having β-ManpA blocks. In contrast, branched α-mannans complexed readily, although the ⁵¹V resonances for one with side chains terminated with α-Manp-(1 \rightarrow 3)-α-Manp-(1 \rightarrow differed from another with only α-Manp-(1 \rightarrow 2)-α-Manp-(1 \rightarrow groups. The anomeric configuration of Me α-Galp and Me β-Galp, each with 3,4-*cis*-diols remote from C-1, gave rise to three ⁵¹V signals of complexes with similar shifts and proportions. The shifts of a galactomannan with terminal α-Galp-(1 \rightarrow 2)-α-Manp- were the same as those with α-Galp-(1 \rightarrow 6)-β-Manp- groups, but fewer complexes were formed with the former structure, probably due to greater steric crowding of the vanadate esters. Most of the complexes gave rise to a signal in the δ –515 region, consistent with the dimeric trigonal-bipyramidal structure.

Keywords: Hexopyranans; VO₃(V) complexes; Vicinal cis-diols; Anomeric configuration; 51V NMR spectroscopy

1. Introduction

show the specificity and degree of complexation in aqueous solution of pentavalent vanadate with polyhydroxylic systems. In 1986, Tracey and Gresser¹ showed that aqueous ethylene glycol and pentavalent Na₃VO₄ formed as a major product, a trigonal–bipyramidal cyclic ester dimer, with a 2:2 stoichiometry, which gave rise to a signal at δ –520. This was followed with a series of other investigations.²⁻⁴ In their review of 1998, Verchére et al.⁵ related that for the previous decade, a controversy existed as to the structure of the main

complexes in aqueous solution, but that the consensus was that they had a distorted trigonal-bipyramidal geometry (1). Clearly, with asymmetric diols, two stereoisomeric complexes can be formed, the simplest system of propylene-1,2-diol giving rise to two signals at δ -523 and -529.1 Tracey and Gresser extended their investigations to cyclic diols and methyl glycosides. The equilibrium constant for complexation of a great excess of 1,2-cyclohexanediols over vanadate with in HEPES buffer at pH 7.5 favored the cis- over the trans-isomer to the extent of 10:1, and 51V NMR spectroscopy of both isomers showed signals at $\delta \sim -520$. In the case of Me β -Manp, with its 2,3-cis-diol, a signal at δ -522 was observed close to one at δ –517 from Me α - and β -Galp. which contain 3,4-cis-diols, but with additional signals at δ -544 and -502. These would be due to complexes with other structures, perhaps related to the 4,6-OH

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groups, perhaps more favorable to complexation than those in the Manp and the α - and β -Glcp series. In the latter, with their *trans*-OH groups in the conformationally stable pyranosyl ring, virtually no complexation took place.

Geraldes and Castro⁷ investigated complexation of reducing monosaccharides with ammonium vanadate at pH 7.0. Glucose gave rise to a signal at δ –523, probably due to complexation with the 1,2-*cis*-diol of its α -anomer.

More recently, Noleto et al.⁸ studied the complexation of NaVO₃ in D₂O at its natural pD, with Me α-Manp, Me α-Galp, and its β-isomer, and selected O-methyl derivatives with defined vicinal cis- and OH-4,6-diols. Me α-Manp and its derivatives having free cis-OH-2,3 groups complexed (the 2,3-complexation site was shown by ¹³C NMR spectroscopy), giving principal ⁵¹V signals in the δ –515 to –517 region. As previously found,⁶ a variety of complexes were formed with Me α-and β-Galp giving signals at $\delta \sim -494$, –508, and –535.⁸ Complexation occurred with Me 6Me- α -Galp and Me 2,6-Me₂- α -Galp, which have 3,4-cis-diols, but not with Me 3Me- β -Galp, which contains free 4,6-OH groups.⁸

As an aqueous solution of a complex of oxovana-dium(IV) with a lichen galactomannan gave rise to activities related to peritoneal macrophages and leish-manicidal activity,⁹ we now examine the formation of esters of selected polysaccharides with NaVO₃ without accompanying buffer, which could be biologically prejudicial. Tested were galactopyranose, mannopyranose-, and mannopyranoseuronic acid-containing polysaccharides, taking into account a surprising preliminary experiment, in which ⁵¹V NMR spectroscopy showed an almost complete lack of complexation with methyl β-mannopyranoside (Table 1).

2. Results and discussion

2.1. Experiments with low-molecular-weight standards

NaVO₃ dissolved in D₂O solution in the presence of carbohydrates, gave rise to 51 V signals of ester complexes at lower field than those of vanadate oligomers, whose signals (areas in parentheses) were at δ –549 (1%), –565 (2%), –570 (78%), and –578 (19%): The latter have been attributed to monomers, dimers, tetramers, and pentamers, respectively. Experiments were now carried out using a 3:1 molar ratio of carbohydrate to NaVO₃, with an equivalent of 194 being used for Me α - and Me β -Manp. The value for polysaccharides corresponded to the mass of each glycosidic unit.

As previously shown, Me α-Manp gave rise to signals of complexes at δ –499 (1%), –506 (2%), and –514 (16%) (Table 1; Fig. 1A),⁸ whereas now the β-anomer was

Table 1. Chemical shifts and percentage areas of ⁵¹V NMR signals of vanadate(V) esters

Substrate	51 V NMR signals of complexes (δ in ppm)
Ethylene glycol, pD 7.5	-515 (3%)
Me α -Man p , pD 7.0^{a}	-492 (5%), -507 (12%), -515 (20%)
Me α -Man p , pD 7.2^{b}	-515 (3%)
Me α -Man p , pD $7.5^{a,8}$	-499 (1%), -506 (2%), -514 (16%)
Me α -Man p , pD 8.0^{a}	-499 (1%), -506 (2%), -515 (17%)
Me α -Man p , pD 8.5^{a}	-499 (1%), -507 (2%), -515 (16%), -541 (2%)
Me α -Man p , pD 9.0^{a}	-499 (1%), -506 (1%), -515 (14%), -533 (6%)
Me α -Man p , pD 10.0^a	-515 (7%), -530 (31%)
S. rouxii α-mannan (2) ^a	-516 (6%)
S. cerevisiae α-mannan (2,3) ^a	-494 (4%), -508 (6%)
S. cerevisiae α-mannan (2,3) ^c	-493 (24%), -508 (35%)
Partly hydrolyzed S. cerevisiae α-mannan ^a	-491 (5%), -517 (5%)
Me β -Man p^a	-496 (1.1%), -506 (0.7%)
<i>R. glutinis</i> $(1 \rightarrow 3), (1 \rightarrow 4)$ - β -mannan $(4)^a$	-518 (0.3%)
L. brasiliensis β-Manp A block (5)(Na ⁺) ^a	-492 (0.3%), -507 (0.5%), -529 (0.1%), -542 (0.2%)
Me α -Gal $p^{a,8}$	-495 (9%), -508 (10%), -535 (4%)
T. fermentans α-galacto-α-mannan (6) ^a	-493 (1%), -508 (1%)
M. scabrella α-galacto-β-mannan (7) ^a	-493 (2%), -508 (3%)
Me β-Gal $p^{a,8}$	-493 (10%), -509 (9%), -534 (3%)

^aMolar equivalent ratio of carbohydrate to NaVO₃, 3:1.

^bA molar equivalent ratio of carbohydrate was used, but the solution was diluted 10-fold.

^cMolar equivalent ratio of carbohydrate to NaVO₃, 12:1.

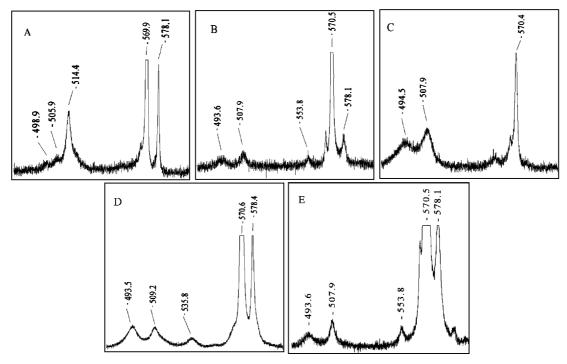


Figure 1. ⁵¹V NMR spectra of D₂O solutions (30 °C) of NaVO₃+Me α-Manp (A), S. cerevisiae α-mannan (mannan to NaVO₃ molar ratio, 3:1) (B), S. cerevisiae α-mannan (mannan to NaVO₃ molar ratio, 12:1) (C), Me β-Galp (D), and galactomannan from M. scabrella (E).

evidenced by small signals at δ –496 (1.1%) and –506 (0.7%) (Table 1). This low level of complexation, referring to structure 1, could be due to steric interaction of the 1-OMe with the vanadate group(s), both being on the β -face of the pyranosyl ring.

Tracey and Gresser¹ showed that ethylene glycol complexed in Tris–Cl buffer at pH 7.5 to give a 2:2 ester dimer with a signal at δ –520. Ray et al.¹⁰ in an aqueous system containing predominantly ethylene glycol showed that a signal at δ –523 was obtained. In our D₂O solution, an isotope effect was observed, since the ester resonance appeared at δ –515 (Table 1).

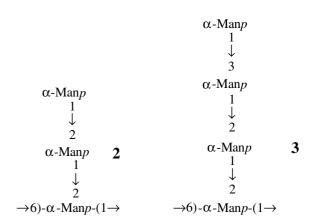
The number and proportions of vanadate esters of Me α -Manp varied according to the pD (Table 1). At all pDs, a signal of a stereoisomeric 2:2 dimer appeared at δ –515. Other minor signals appeared at –499 and –506, which were also not pD dependent. At pD 8.5–10.0, a signal at higher fields became increasingly prominent, finally appearing at δ –533 (31%).

An aqueous solution of NaVO₃, at the same concentration as used in ^{51}V experiments, had pH 7.49. It changed very little to pH 7.55 on addition of a 3:1 amount of Me α -Manp.

2.2. Complexation of vanadate with branched α-mannans

An α -mannan from *Saccharomyces rouxii* contains structure **2**, and another with one less nonreducing endunit, both containing 2,3-*cis*-diols. ^{11,12} Each, with its terminal α -Manp-(1 \rightarrow 2)-unit, gave rise to a single ⁵¹V NMR signal at δ –516 (Table 1).

The α -mannan from Saccharomyces cerevisiae (baker's yeast), with predominant repeating structure 3^{13} having terminal α -Manp- $(1 \rightarrow 3)$ - and some α -Manp- $(1 \rightarrow 2)$ -units, as in **2**, gave rise to two signals at δ –494 (4%) and –508 (6%) (Table 1, Fig. 1B). On increasing the α -mannan to vanadate ratio by a factor of 4 to 12:1, the resulting 51 V spectrum, as expected, showed greatly enlarged ester signals at δ –494.5 (24%) and –508 (35%) (Table 1; Fig. 1C).



Partial hydrolysis of *S. cerevisiae* α -mannan preferentially removed its side chains ¹⁴ to form a product with 41% of $(1\rightarrow 6)$ -linkages, as determined by the relative area of its H-1 signal at δ 5.16, which resonates at highest field of all its H-1 signals. However, even with more nonreducing end units than those of the parent α -mannan, no appreciable increase in complexation took

place with signals at δ –491 (5%) and –507 (5%) (Table 1)

An attempt was made to determine the complexation sites in the α -mannan using ^{13}C NMR spectroscopy, despite the broadness of its signals when compared with those of Me α -Manp. However, two possible signals were detected at δ 87.1 and 89.7, which when compared with those of esters of Me α -Manp, 8 at δ 87.4 (C-2) and 80.0 (C-3), suggested that the former could arise via esterification at C-2. However, any accompanying C-3 signal would overlap with those of 2-O- and 3-O-substituted units of the α -mannan, which would appear in the δ 80.0 region. 15

2.3. Complexation with linear β -mannan and β -mannopyrananuronic acid

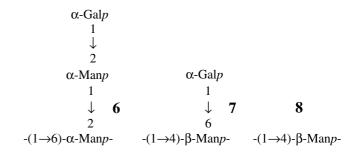
The low degree of complexation found with Me β -Manp also extended to β -linked polysaccharides. Two with free *cis*-2,3-diols were examined. One was from *Rhodotorula glutinis* (4),¹⁶ which formed only a very small signal of complex at δ –518 (0.3%) (Table 1).

$$[-\beta$$
-Man p -(1 \rightarrow 3)- β -Man p -(1 \rightarrow 4)- $]$ n 4
 $[-\beta$ -Man p A-(1 \rightarrow 4)- $]$ n 5

The other was the sodium salt of a $(1 \rightarrow 4)$ -linked β -mannopyranosyluronic acid block (5), isolated from alginic acid of the marine alga *Laminaria brasiliensis*. ¹⁷ This also barely complexed, as shown by small signals at δ -492 (0.3%), -507 (0.5%), -529 (0.1%), and -542 (0.2%) (Table 1).

2.4. Complexation of vanadate with $\alpha\text{-galactopyrano-}\alpha\text{-mannan}$ and $\alpha\text{-galactopyrano-}\beta\text{-mannan}$

The degrees of complexation of vanadate with an α galacto-α-mannan and an α-galacto-β-mannan, each having terminal α-galactopyranosyl units, were determined. It has been shown that there was no anomeric effect influencing its complexation with Me α-Galp and Me β-Galp, each giving rise to three signals with similar chemical shifts and proportions (Table 1),8 with only those of Me β-Galp being presented in Table 1 and Figure 1D. A galactomannan of the yeast *Trichosporon* fermentans¹⁸ should complex with vanadate by virtue of its Me α-Galp terminal units present in repeating structure 6. However, although two similar resonances were present at -493 (1%) and -508 (1%), these were much smaller that those of the Me α -Galp standard, which are at δ -495 (9%), -508 (10%), and -535 (4%) (Table 1).8



A somewhat greater degree of complexation, although smaller than that observed with Me α -Galp, was found with the galactomannan from seeds of *Mimosa scabrella*, which contains structures **7** and **8**^{19,20} (it is assumed that the 4-O- and 4,6-di-O-substituted β -Manp units of the main chain would not contribute to the vanadate ester signals). When compared with signals of structure **6**, similar ones of complexes were formed, but in a greater proportion at δ –494 (3%) and –508 (3%) (Table 1; Fig. 1E). It is interesting that the high field of the methyl galactosides does not appear in the spectra of the two galactomannans. This point will be further investigated.

In both galactomannans, the low proportion of signals of the complexes could be due to nonbonded hindrance of the adjacent α -Manp unit, which would crowd the vanadate ester of the 3,4-cis-diol. It would be expected that structure 6, which has \rightarrow 2)- α -Manp-, rather than the less sterically hindering \rightarrow 6)- α -Manp-units in 7, would give rise to a greater free energy and less complexation.

3. Conclusions

A number of mannans, galactomannans, and a β-mannopyrananuronic acid, containing vicinal cis-diol groups, were treated with in D₂O solution with NaVO₃ $(pD \sim 7.5)$ to form complexes of vanadate ester, detected and quantified by 51V NMR spectroscopy. Mannosecontaining polysaccharides with 2,3-cis-diols in the α-Manp series and galactomannans with 3,4-cis-diols in the α - and β -Galp series complexed, as evidenced by low-field signals of their esters. Much less complexation occurred with a β -mannopyranan and a β -mannopyrananuronic acid, each containing 2,3-cis-diols, unlike polysaccharides containing α -Manp units, due to steric effects. The greatest degree of complexation occurred with the branched α-mannan from S. cerevisiae, followed by the branched galactomannan of M. scabrella. The complexes of these polysaccharides will be tested as targeting devices to see if they have leishmanicidal activity.

4. Experimental

4.1. Materials

Sodium metavanadate (NaVO₃) was obtained from Sigma-Aldrich, as was methyl β -D-mannopyranoside, which was labeled in the catalogue as its *iso*-propylate. Its ^{13}C NMR spectrum showed that it was the glycoside with 2-propanol of crystallization. To remove this, it was dissolved in H₂O and freeze dried, and ^{13}C NMR examination of the residue showed it to be identical with authentic material. Other carbohydrates were available in our laboratory.

4.2. pH effect on addition of Me α -Manp to vanadate solutions

As an indication of pD values used in 51 V examination of polysaccharides, a solution of vanadate (110 mg) in water (4 mL) had pH 7.49. On addition of Me α -Manp (450 mg), the pH was 7.55. At a 10-fold dilution of the above, the pH was 7.20, before and after addition of Me α -Manp.

4.3. Preparation of solutions of carbohydrate with sodium vanadate

Aqueous solutions of polysaccharide (500 mg in 10 mL) were adjusted, if necessary, to pH 7.5, freeze-dried, and 45 mg of the residues were dissolved in D_2O (0.5 mL). To this solution, sodium vanadate (11 mg) in D_2O (0.4 mL) was added dropwise slowly with shaking, in order to avoid formation of clots of complex.

In the calculation of millimolar concentrations, a molar equivalent of 194 was used for Me α -Manp and Me β -Manp, 162 for hexosans, 198 for the alginate (Na⁺ salt), and 62 for ethylene glycol. In general, 3:1 carbohydrate to NaVO₃ molar ratios were used, although one of 12:1 ratio was also examined with the α -mannan of S. cerevisiae. The galactomannan of M. scabrella, which formed viscous solutions, was dissolved in 3 mL of D₂O. In general, the solution of vanadate was mixed with that of polysaccharide solution with vigorous shaking, in order to avoid possible clots of vanadate complex.

4.4. NMR spectroscopy

NMR measurements were carried out in 5-mm tubes, on D₂O solutions at 30 °C, using a Bruker DRX-400 Avance spectrometer. ¹³C spectra were obtained at 100.6 MHz with 1 s delay and 0.6 s acquisition times, and 80,000 transients in attempts to detect minor signals of polysaccharide complexes (this was only successful with the *S. cerevisiae* α -mannan). Chemical shifts are expressed as δ (ppm) relative to that of external Me₄Si (δ = 0).

The ¹H spectrum which was of acid-degraded *S. cerevisiae* mannan was obtained at 30 °C at 400.16 MHz with presaturation with 2 s delay and acquisition time and 16 transients.

 51 V spectra were obtained at 105.2 Hz, with 0.5 s delay and 0.10 s acquisition times, and 1000 transients. Chemical shifts are in δ (ppm) relative to that of external VOCl₃ (δ = 0).

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